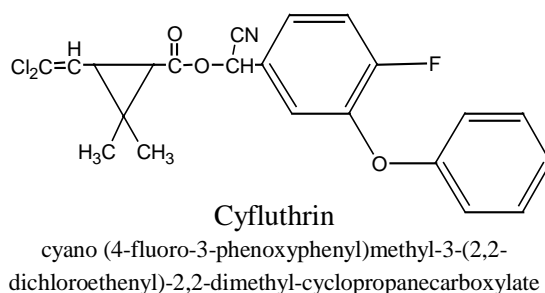


Environmental Fate of Cyfluthrin

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This document reviews the environmental fate of cyfluthrin, chemical name Cyano(4-fluoro-3-phenoxy-phenyl)methyl-3-(2,2-dichloroethenyl)-2,2-dimethylcyclopropanecarboxylate. Products containing the active ingredient cyfluthrin include Attatox, Bay FCR 1272, Baythroid, Baythroid H, Contur, Laser, Solfac, and Tempo among many others.



General Information and Mode of Action

Cyfluthrin is a nonsystemic synthetic pyrethroid insecticide used to control chewing and sucking insects. Through contact and stomach poisoning, it attacks the nervous system; resulting in swift debilitation and has a long residual effect. Insects that come into contact with cyfluthrin die of starvation and desiccation due to being weakened by the chemical and thus, ceasing to feed. Its uses are extensive, including: agricultural crops, stored products, public health situations (i.e. cockroaches, mosquitoes, and flies), ornamentals, turf, and domestic pests. Target insects include ants, silverfish, cockroaches, grain beetles, fleas, flies, European corn borer, Colorado potato beetle, and many others (Farm Chemical Handbook, 1993; EXTOWNET, 2001; Hamman and Fuchs, 1981).

Cyfluthrin is a clear liquid, dark amber in color, with an oily to pasty consistency, and has a faint aromatic solvent odor at room temperature. It is thermally stable at room temperature. Four diastereoisomeric pairs of enantiomers make up cyfluthrin:

- I (R)- α -cyano-4-fluoro-3-phenoxybenzyl (1R)-cis-3-(2,2-dichlorovinyl)-2,2-dimethylcyclopropanecarboxylate + (S)- α , (1S)-cis-;
- II (S)- α , (1R)-cis + (R)- α , (1S)-cis-;
- III (R)- α , (1R)-trans- + (S)- α , (1S)-trans-; and
- IV (S)- α , (1R)-trans- + (R)- α , (1S)-trans- (Tomlin, 1997; Hamman and Fuchs, 1981).

Physical-Chemical Properties^a

Molecular weight	434.29
Water solubility	2.0×10^{-3} ppm (20°C)
Vapor pressure	3.24×10^{-8} mm Hg (25°C)
Hydrolysis half-life	193 days (25°C at pH 7)
Aqueous photolysis half-life	12.2 days (25°C at pH 5)
Anaerobic half-life	33.6 days (loam)
Aerobic half-life	63 days (25°C at pH 5.9; sandy loam)
Soil photolysis half-life	2-16 days (28°C at pH 6.6; sandy loam)
Field dissipation half-life	~ 13.5 days (pH 5.8; sandy loam)
Henry's law constant	4.93×10^{-6} atm m ³ /mole (20°C)
Octanol-water coefficient (K _{OW})	4.58×10^5 - 6.4×10^5
Soil adsorption coefficient:	
K _D	9.4 - 28
K _{OC}	6.24×10^4

Toxicity^b

<i>Daphnia magna</i> LC ₅₀ , 48 hr	0.16 ppb
Mysid shrimp LC ₅₀ , 96 hr	0.0025 ppb
Rainbow trout LC ₅₀ , 96 hr	0.302 ppb
Bluegill Sunfish LC ₅₀ , 48 hr	0.998 ppb
Fathead Minnow LC ₅₀ , 96 hr	2.49 ppb
Bobwhite quail LD ₅₀ , 8 days	>5,000 ppm
Mallard duck LD ₅₀ , 8 days	>5,000 ppm
Rat acute oral LD ₅₀ , Female	1271 ppm
Rat acute oral LD ₅₀ , Male	869 ppm

^a DPR Pesticide Chemistry database (2001)

^b DPR Ecotox database (2001)

Environmental Fate

Air

When applied aerially or by broadcast spray, there is possibility for drift. However, cyfluthrin is relatively non-volatile, as indicated by its low vapor pressure (3.24×10^{-8} mm Hg) and low Henry's law constant (4.93×10^{-6} atm m³/mole). In addition, the strong tendency to partition to soil reduces the potential for volatilization as a major route of dissipation.

Water

With low water solubility and a high K_{OC} , cyfluthrin has a strong tendency to absorb to soil. Although cyfluthrin may not be readily degraded in water at a neutral pH, the aqueous hydrolysis half-life decreases with an increase in pH. Anderson (1986) reports a hydrolysis half-life in water of approximately 4 days at pH 8.3-8.6, with a significant slowing of degradation after one week. At pH 9, the hydrolysis half-life is 2 days (DPR Pesticide Chemistry database, 2001). A relatively short aqueous photolysis half-life of 12.2 days at pH 7 suggests that an important route for the dissipation of cyfluthrin is through aqueous photolysis.

In a study by Heimbach et al. (1992), concentrations of cyfluthrin decreased rapidly, with maximum levels between 4 hours to 1 day post application in artificial ponds. In natural ponds, maximum levels were detected between 1 and 2 days post application and then rapidly declined. Cyfluthrin was not detectable at 2 days post application in either the artificial or natural ponds.

Cyfluthrin degrades abiotically through hydrolysis. The major degradation product is 4-fluoro-3-phenoxybenzoic acid (FPB acid), which is formed by the first breakdown products; permethric acid and the cyanhydrin of 4-fluoro-3-phenoxybenzaldehyde (FPB ald). Other degradation products include FPB ald, 4'-OH-FPB acid, 'amide'-baythroid (2-amino-1-(4-fluoro-3-phenoxyphenyl)-2-oxoethyl 3-(2,2-dichloroethenyl)-2,2-dimethylcyclopropanecarboxylate), and "acid"-baythroid (α -[[[3-(2,2-dichloroethynyl)-2,2-dimethylcyclopropyl]carbonyl]oxy]-4-fluoro-3-phenoxybenzenoic acid (Anderson, 1986; Preiss et al, 1984).

Soil

Cyfluthrin is hydrophobic and has a high K_{OC} . Consequently cyfluthrin has a low tendency to leach in soil. In addition, cyfluthrin mobility during runoff events will be low, except under conditions of high sediment transport. Under these conditions, cyfluthrin may move off-site with suspended sediment. Lin (1992) reported that 0.3 % of the initial application moved off-site during a rain event following application to cotton in a fine sandy loam. At another site, Lin (1992) found that 0.21% of total application moved off-site after 4 rain events. Shehata et al. (1987) reports that the concentration of applied cyfluthrin or the volume of leaching water did not affect cyfluthrin's binding behavior or leachability.

As with most pyrethroids, the persistence of cyfluthrin is highly variable due to various environmental conditions. In example, the persistence of cyfluthrin in soil was found to be unaffected by soil moisture but affected by organic matter and redox potential (Smith et al., 1995). Degradation of cyfluthrin was more rapid at higher pHs, thus, cyfluthrin was observed to be more persistent in acidic soils than in alkaline soils (Shehata et al., 1987).

In various field dissipation studies, cyfluthrin showed low persistence; its field dissipation half-life is approximately 13.5 days at pH 5.8 in sandy loam soil (DPR Pesticide chemistry database, 2001). Stout and Leidy (2000) report that the mean residues of microencapsulated cyfluthrin declined from maximal levels after three days, but residues were detectable through day 45. In a rotational crop field study, cyfluthrin was found at 0.36 ppm residue in silty soil and ranging from 0.06-0.24 ppm for clay soils on the day of application. At planting and harvest (day 135 and 330 post application), the total residue in clay was detected at less than 0.01 ppm (Leslie, 1988). In a study conducted on loam and sandy loam soils, the dissipation half-life of cyfluthrin ranged from 3.9 days to 23.8 days. At day 29 post-application, no measurable residues were found. By day 60, the FPB acid and Cl₂Ca (3-(2,2-dichlorovinyl)-2,2-dimethyl cyclopropane carboxylic acid) metabolites had dissipated to less than 0.01 ppm (Grace and Cain, 1990), suggesting that they are also not persistent. Wagner et al. (1983) also observed rapid aerobic degradation in both loam and sandy loam soils with half-lives of 56 and 63 days, respectively. The major degradation products found in cyfluthrin were: 4-fluoro-3-phenoxybenzaldehyde (FPB ald), 4-fluoro-3-phenoxy benzoic acid (FPBacid), Cl₂Ca and 'amide'-Baythroid. Trace amounts of 'acid'-Baythroid, 4-fluoro-3-phenoxybenzenemethanol (FPB alc), and 4'-OH-FPBacid were also detected in several studies. Cyfluthrin completely degrades with mineralization to CO₂ (Grace and Cain, 1990; Gronberg and Pfankuche, 1983; Wagner et al., 1983;)

Biota

Vegetation - A nonsystemic insecticide, cyfluthrin has a low tendency to penetrate into plant tissues and the translocation of cyfluthrin appears negligible, with very low concentrations found in other parts of the plants (Tomlin, 1997). In a three-year replicated study, no significant difference was found in photosynthesis, stomatal conductance, or transpiration rates of alfalfa or soybean due to cyfluthrin. However, the rate of photosynthesis of alfalfa increased one hour post application in one replicate year, suggesting a possible, immediate, but short-lived, increase in the rate of photosynthesis in alfalfa (Haile et al., 1999). The major metabolites of cyfluthrin in plants are 3-phenoxy-4-fluorobenzylalcohol and FPB acid (Preiss et al., 1984).

In a dislodgeable residue study, rapid dissipation was reported. On applications of cyfluthrin to cotton leaves, dislodgeable residues ranged from 0.0217 µg/cm² 8-10 hours post-application. Residues declined to 0.0007 µg/cm² 27 days post-application (Loeffler, 1986).

Animals and Fish – Following an experimental application (for assessment of hazards to aquatic ecosystems) of 12.5 g cyfluthrin/hectare to artificial ponds, there were no

mortalities in trout nor did it affect their development (Heimbach et al., 1992). An exposure of 0.13 ppb cyfluthrin to bluegill sunfish resulted in mean fish tissue residues being greater than 20 ppb at day 0 to 63 ppb day 28. The bioconcentration factor reached a maximum of 854 on day 14 (Carlisle and Roney, 1984). Although extremely toxic to fish and other aquatic organisms, cyfluthrin is relatively non-toxic to upland game birds and water fowl. It is also toxic to the honeybee, LC₅₀ of 0.037 mg/bee. Cyfluthrin was found to be eliminated quickly in mammals, with 98% of the administered amount being excreted via urine and feces after 48 hours (Tomlin, 1997; EPA Fact Sheet, 1987).

Soil Biota - Tu (1995) reported an increase in bacterial populations during 2 weeks post application but observed no effects on fungal populations, oxidation of soil, or levels of nitrification or denitrification. It was concluded that soil indigenous microbes were able to tolerate cyfluthrin when it was applied to control soil pests.

Summary

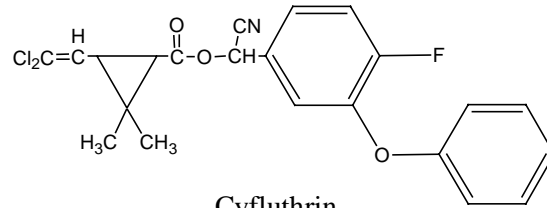
A nonsystemic synthetic pyrethroid, cyfluthrin is used in the control of chewing and sucking insects. It poisons the stomach and attacks the nervous system, weakening the insect and thus, starving it. Although primarily used in agriculture, cyfluthrin is also used in public health situations, and for domestic pests.

A low vapor pressure of 3.24×10^{-8} mm Hg indicates that cyfluthrin is non-volatile in air while a low Henry's law constant of 4.93×10^{-6} atm m³/mole indicates a low volatility in water. A high K_{OW} ranging from 4.58×10^5 - 6.4×10^5 suggests a strong tendency to partition from water to soil. In various soils, cyfluthrin has been observed to degrade rapidly and has shown little leaching potential. The hydrolysis half-life of 193 days at pH 7 suggests that cyfluthrin is not readily degraded in water; however, its degradation will increase with an increase in pH. In vegetation, cyfluthrin does not penetrate significantly into plant tissues, and does not appear to affect photosynthesis, stomatal conductance, or transpiration rates. Cyfluthrin is extremely toxic to fish, other aquatic organisms, and honeybees, but is relatively non-toxic to mammals.

Although cyfluthrin displays high aquatic toxicity in laboratory studies, the tendency to sorb strongly to suspended sediment and dissolved organic materials in field aquatic systems probably reduces cyfluthrin's bioavailability, hence cyfluthrin's aquatic toxicity. However, the extent to which bioavailability is mitigated, and the aquatic toxicity of a hydrophobic pyrethroid is reduced in the water column or in sediments is uncertain, and remains a subject of on-going study and debate (EPA SAP, 1999)

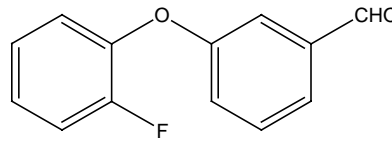
Cyfluthrin Degradation

Photolysis in water

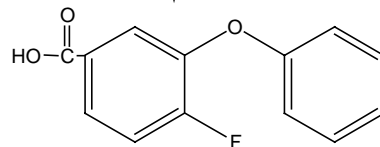
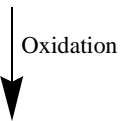


Cyfluthrin

cyano (4-fluoro-3-phenoxyphenyl)methyl-3-(2,2-dichloroethenyl)-2,2-dimethyl-cyclopropanecarboxylate

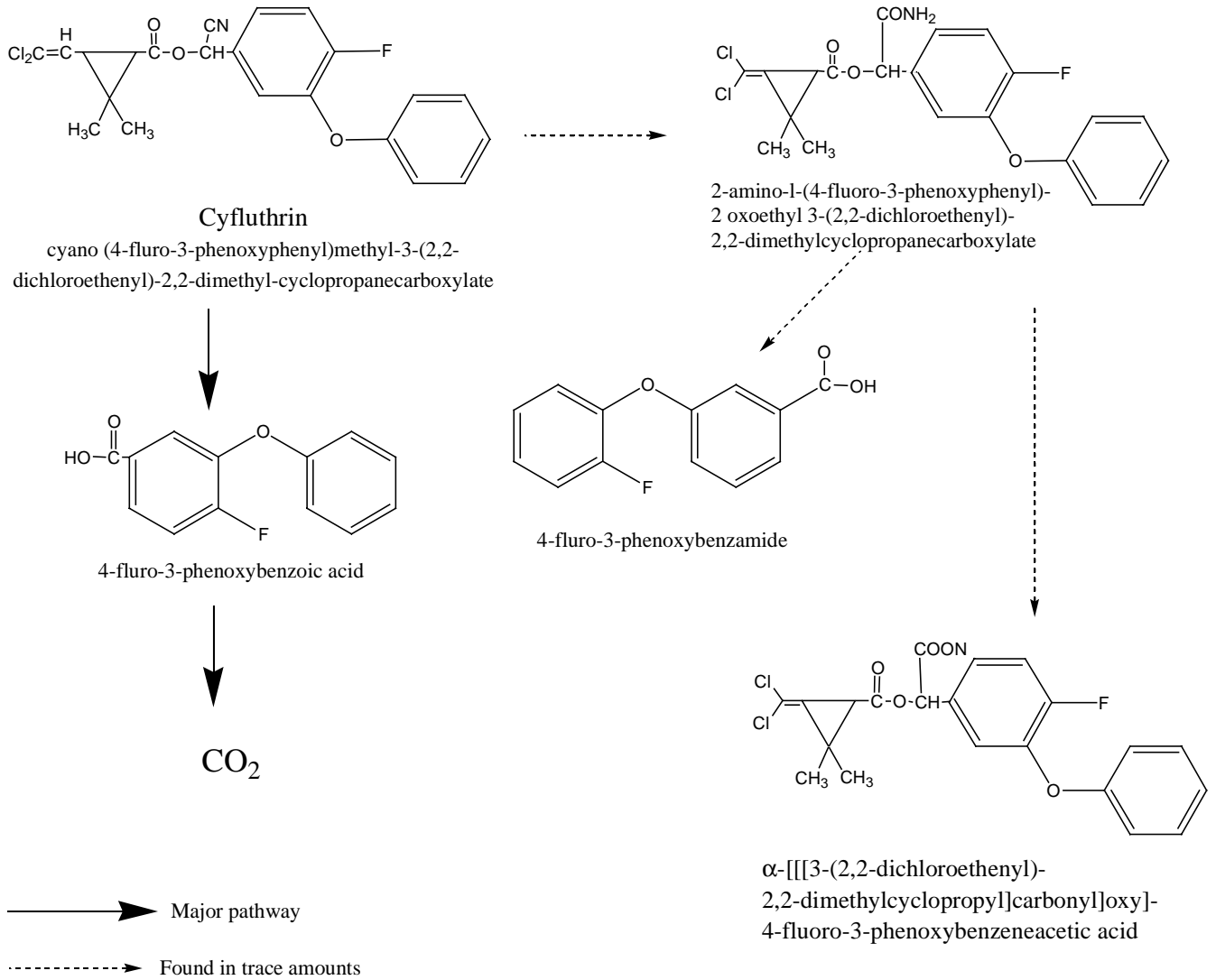


4-fluoro-3-phenoxybenzaldehyde

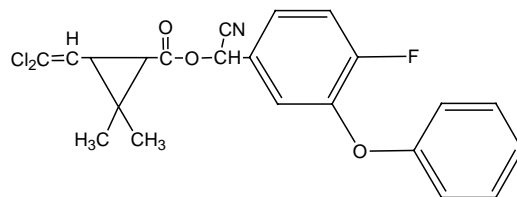


4-fluoro-3-phenoxybenzoic acid

Soil Degradation – Aerobic

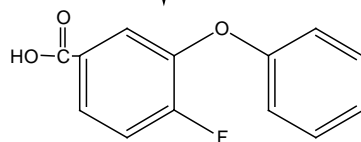


Soil Degradation – Anaerobic



Cyfluthrin

cyano (4-fluoro-3-phenoxyphenyl)methyl-3-(2,2-dichloroethenyl)-2,2-dimethyl-cyclopropanecarboxylate



4-fluoro-3-phenoxybenzoic acid

References

- Anderson, C. 1986. Degradation behavior of ^{14}C -Cyfluthrin in natural water. Mobay Chemical Corp. Report No. 93044
- Carlisle, J. and D. Roney. 1984. Bioconcentration of cyfluthrin (Baythroid) by bluegill sunfish. Mobay Chemical Corp. Report No. 86215
- DPR Ecotox database. Environmental Monitoring Branch, Department of Pesticide Regulation. 2001
- DPR Pesticide Chemistry database. Environmental Monitoring Branch, Department of Pesticide Regulation. 2001
- EPA Pesticide Fact Sheet: Cyfluthrin. 1987. Fact Sheet No. 164
- EXTOXNET. 2001. Cyfluthrin. Available online
<http://ace.orst.edu/cgi-bin/mfs/01/pips/cyfluthr.htm>
- Farm Chemical Handbook. 1993. Meister Publishing Company. Vol. 79
- Grace, T. and K. Cain. 1990. Dissipation of cyfluthrin in California soils. Mobay Chemical Corp. Report No. 100152
- Gronberg, R. and Pfankuche L. 1983. An analytical residue method for BaythroidTM and its major metabolites in soil. Mobay Chemical Corp. Report No. 85886
- Haile, F., R. Peterson, and L. Higley. 1999. Gas-exchange responses of alfalfa and soybean treated with insecticides. *J of Economic Entomology*, 92(4): 954-959
- Hamman, I. And R. Fuchs. 1981. [®]Baythroid, a new insecticide. *Pflanzenschutz-Nachrichten*, 34(2):121-151
- Heimbach, F., W. Pflueger, and Ratte, H. 1992. Use of small artificial ponds for assessment of hazards to aquatic ecosystems. *Environmental Toxicology and Chemistry*, 11:27-34
- Leslie, W. 1998. Baythroid[®] - Residues in field rotational cereal crops. Mobay Chemical Corp. Report No. 98429
- Lin, J. 1992. Field measurements of cyfluthrin fate and runoff from a cotton field in Benoit, Mississippi. Miles, Inc., Report No. 103918
- Lin, J. 1993. Field measurements of cyfluthrin fate and runoff from a cotton field in Southern Georgia. Miles, Inc., Report No. 103919

- Loeffler, W. 1986. ®Baythroid dislodgeable residues on cotton leaves. Mobay Chemical Corp. Report No. 91032
- Preiss, U., K. Wagner, L. Oehlmann, G. Engelhardt, and P. Wallnöfer. 1984. Metabolism of the insecticide Baythroid by cell suspension cultures. *Chemosphere*, 13(8):861-872
- Shehata, A., Abdell-All, A. Farag, M. Samaan, and Y. Mohareb. 1987. Some factors influencing persistence, hydrolysis, downward movement and leachability of cyfluthrin in Egyptian soils. *Annals Agricultural Science*, 32(3):1677-1687
- Smith, S., G. Willis, and C. Copper. 1995. Cyfluthrin persistence in soil as affected by moisture, organic matter, and redox potential. *Bull Environ Contam Toxicol*, 55:142-148
- Stout, D. and R. Leidy. 2000. A preliminary examination of the translocation of microencapsulated cyfluthrin following applications to the perimeter of residential dwellings. *J of Environ Sci Health*, B35(4):477-489
- Tomlin, C, Ed. 1997. *The Pesticide Manual*. British Crop Protection Council. 11th Ed.
- Tu, C. 1995. Effect of five insecticides on microbial and enzymatic activities in sandy soil. *J of Environ Sci Health*, B30(3):289-306
- EPA Scientific Advisory Panel (SAP). 1999. US EPA Scientific Advisory Panel (SAP): Environmental Fate Assessment for the Synthetic Pyrethroids. Available online: <http://www.epa.gov/scipoly/sap/1999/index.htm>
- Wagner, K., H. Neitzel, and L. Oehlmann. 1983. Decomposition of BaythroidTM in soil under aerobic and anaerobic conditions. Mobay Chemical Corporation, Report No. 86052